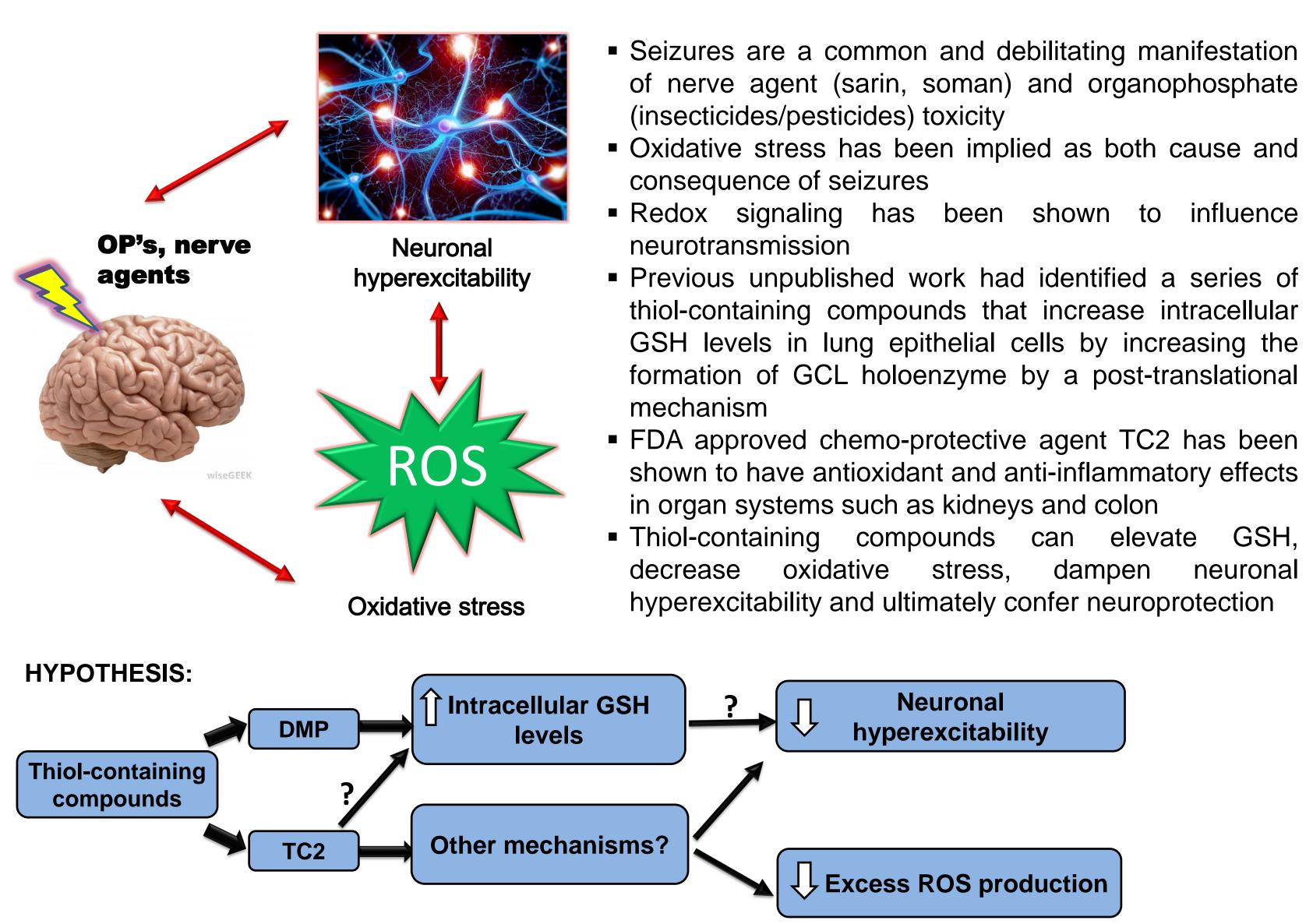


THIOL-CONTAINING COMPOUNDS ATTENUATE OXIDATIVE STRESS AND NEURONAL HYPEREXCITABILITY IN VITRO

ABSTRACT

Chemical agents such as industrial chemicals, pesticides, and chemical warfare agents can induce uncontrolled seizure activity (neuronal hyperexcitability). Oxidative stress has been implicated as a pathogenic factor in the etiology of seizures and epilepsy. However, whether and how **cellular redox status** modulates **neuronal** hyperexcitability is unclear. We hypothesized that the modulation of cellular redox status with thiol-containing compounds would decrease oxidative stress, and attenuate neuronal hyperexcitability in vitro. 2,3-dimercapto-1propanol (DMP), a thiol-containing compound significantly (p<0.001 vs vehicle control) increased intracellular glutathione levels in mixed rat primary cortical cultures at 4 and 24h. Next, we determined if DMP could dampen "seizure-like" activity in vitro induced by 4-Aminopyridine (4AP), a toxicant that inhibits potassium channels. In mixed rat primary neuronal-glial cultures, incubation with 100µM DMP for 4h significantly (p<0.0001 vs 1mM 4AP) decreased 4AP-induced neuronal hyperexcitability. We tested the ability of another thiol compound, TC2 which is FDA approved as a systemic protective agent against chemotherapy, for its ability to alter oxidative stress and neuronal hyperexcitability in vitro. To visualize and quantify the oxidative stress response in primary cortical cultures, we utilized a highly sensitive fluorescent probe, HKSOX-1r, that could detect endogenous superoxide levels. Co-treatment with TC2 decreased Antimycin A-induced superoxide levels to control values. In addition, 500µM TC2 significantly (p<0.0001 vs 1mM 4AP) attenuated 4AP induced neuronal hyperexcitability in mixed rat primary cortical cultures. Taken together, the data suggest that thiol-containing compounds decrease oxidative stress and attenuate neuronal hyperexcitability.

BACKGROUND



METHODS

- * HPLC measurement of intracellular GSH and GSSG levels: Intracellular GSH and GS in BV2 cell line and primary cortical culture cells by HPLC with electrochemical detection modifications to the method described previously (Lakritz et al., 1997 and Beal et al., 1990).
- * PAGE: For the native PAGE, BV2 (murine microglial cell line) cell lysates were probed with antisera (1:10,000). For SDS-PAGE, the lysates were probed with GCLC (1:10,000) ar antibodies.
- HPLC measurement of GCL activity: GCL activity was measured in BV2 cells using method previously described by Gregg et al., 2002.
- Confocal imaging of primary cortical cultures: Endogenous superoxide levels in cultures were imaged using a specialized probe (1r – courtesy of Prof. Dan Yang). Fluore were done using Image J.
- Microelectrode Array (MEA) method: Neuronal excitability measurements in mixed rate were performed using the Maestro MEA system (Axion Biosystems). Briefly, cells were pla MEA plates that can detect neuronal electrical activity.

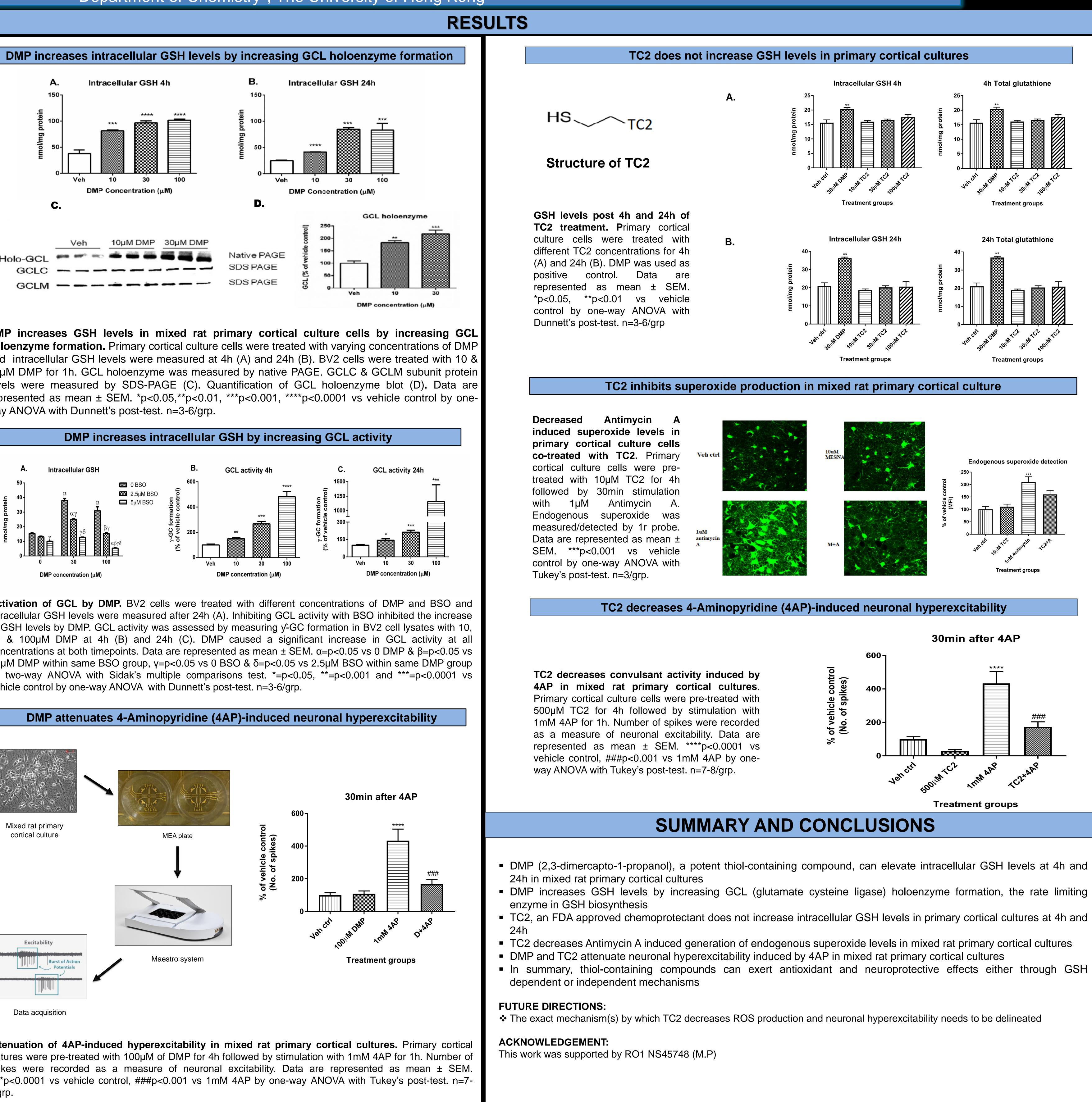
RESULTS

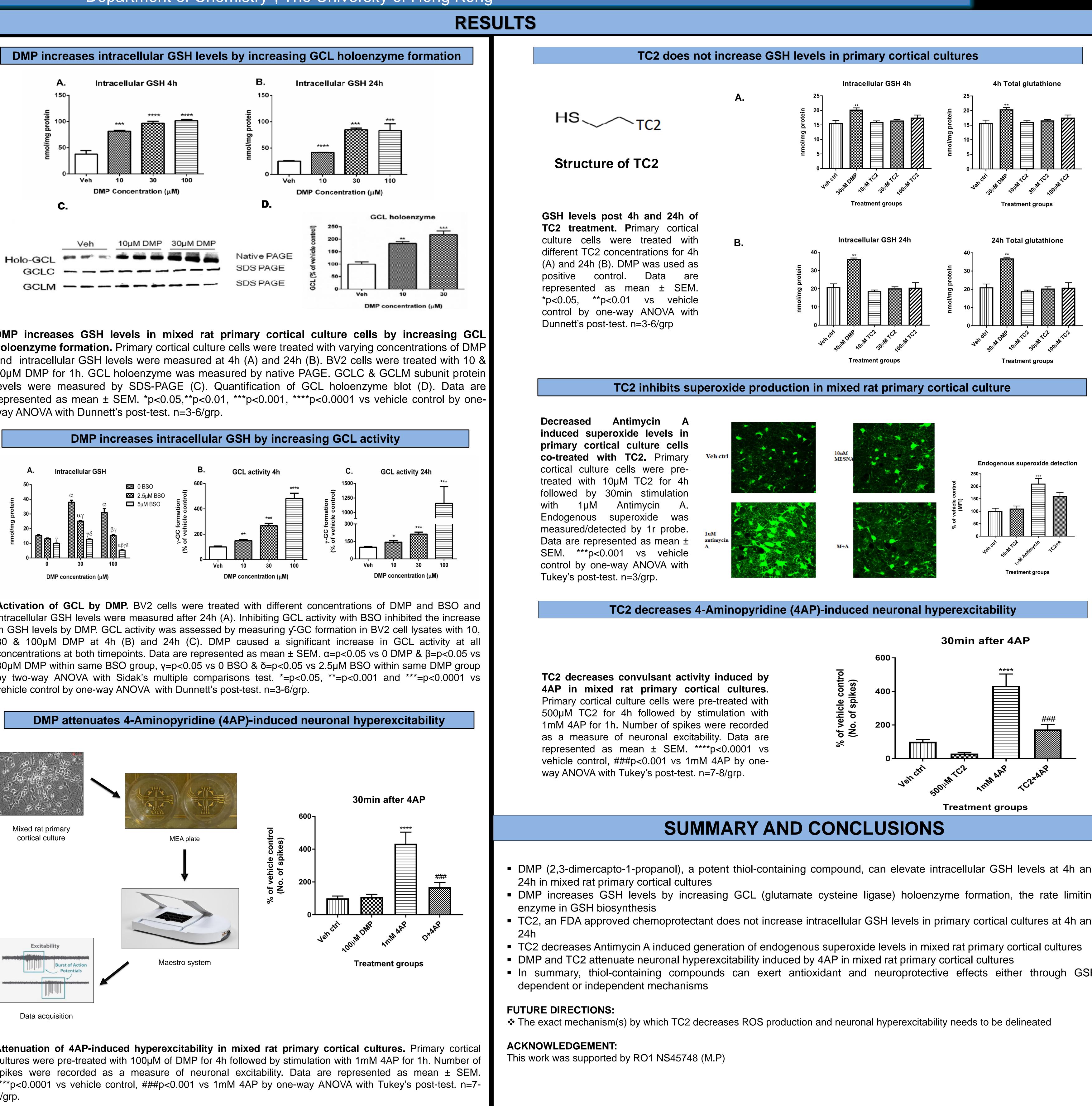
Induction of intracellular GSH (% of vehicle control)				
Thiol compound	Structure	10µM	30µM	100µM
2,3-Dimercapto-1- propanol (DMP)	HS OH SH	147 ± 3**	162 ± 6***	150 ± 14***

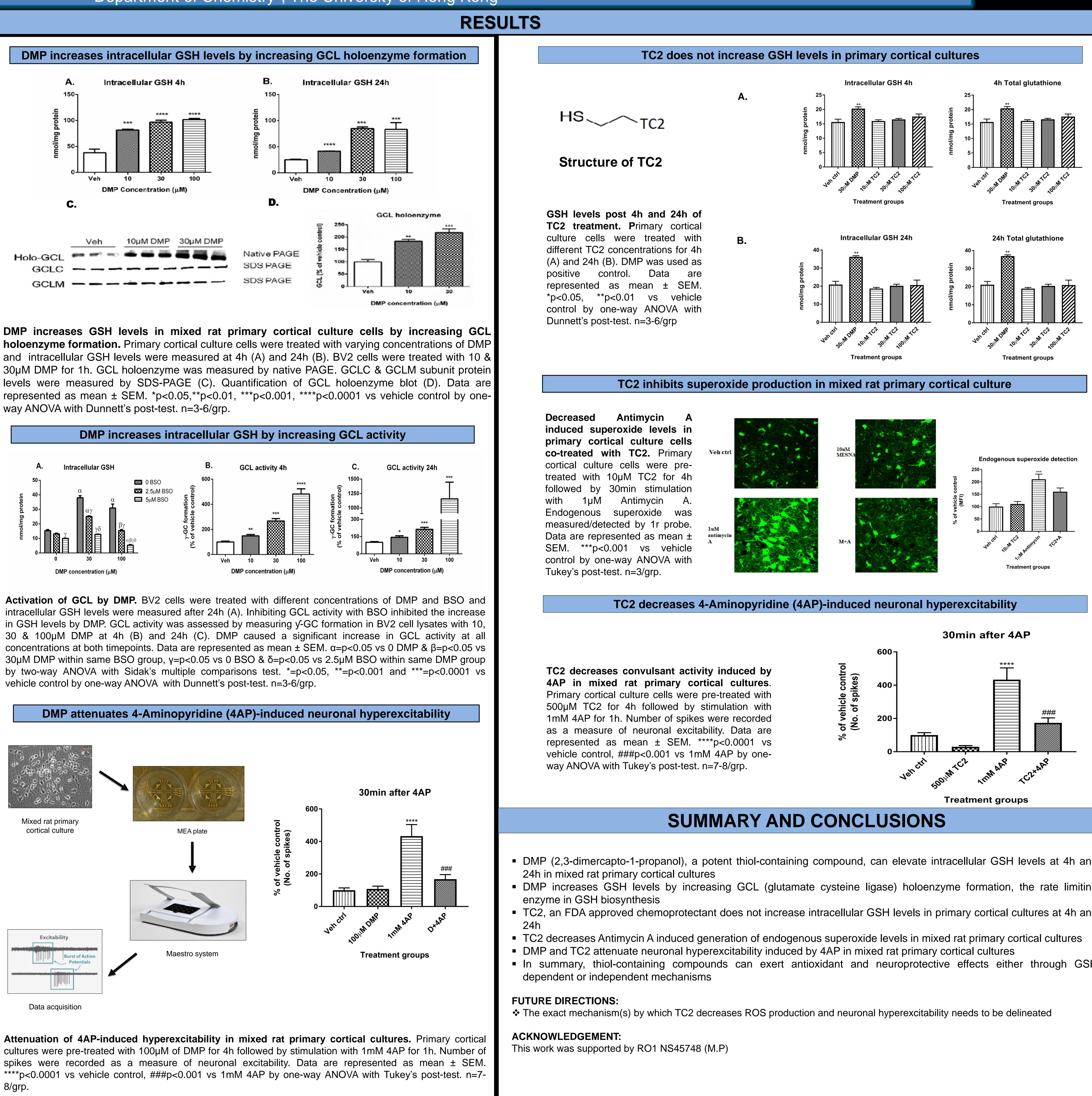
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Intracellular GSH 4h







elevate GSH, oxidative stress, dampen neuronal

ity
duction
SSG levels were measured (HPLC-EC) following minor
vith polyclonal GCLC rabbit and GCLM (1:5000) rabbit
ng HPLC-EC following the
mixed rat primary cortical escence intensity analyses
at primary cortical cultures ated and treated in 48-well
ion of intracellular GSH IP. BV2 cells were treated 0, 30 and 100µM of DMP tracellular GSH levels were red at 4 hours. Data are

ented as mean ± SEIM and ***p<0.001 vs vehicle by one-way ANOVA with tt's post-test. n= 3-6/grp.

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